

Understanding the impact of IFN γ R clustering on immune response pathways

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Short Abstract — Interferon- γ plays an important role in macrophage activation during the early steps of innate immunity. Propagation of immune response via IFN γ is dependent on the spatial localization of the IFN γ R. EM images have found IFN γ R to be colocalized in caveolar membrane domains; whether this enhances or restricts signal remains to be elucidated. However, in an interesting twist, experimental evidence points to IFN γ as a negative regulator of caveolin-1. In order to understand the spatial-temporal dynamics of IFN γ R membrane localization and further investigate the impact of IFN γ R activation on gene and metabolic pathways that regulate caveolin-1 production we developed a simulation-based model using a coupled CSNSA-BioXyce platform that combines a spatial Monte Carlo method (CSNSA) with a circuit-based intracellular network simulator (BioXyce). In this work we explore the impact of receptor spatial organization on immune effector mechanisms and to complete the circle, the impact of IFN γ mediated effectors on spatial organization.

Keywords — Spatial organization, gene networks, IFN γ R, caveolin-1, spatial modeling

I. PURPOSE

Interferon- γ induces direct antimicrobial mechanisms and up-regulates antigen processing and presentation pathways (9). Interferon- γ activates these immune responses via the Janus kinase (JAK) and signal transducer and activator of transcription (STAT) pathway. The initial signaling events of IFN γ receptor (IFN γ R) are often the rate limiting step and dependent on the spatial distribution of the IFN γ receptors (3).

Electron microscopy using immunogold labeled particles has revealed aggregation of IFN γ R in membrane domains (1). With low numbers of receptors (10^2 to 10^3 receptors per cell (1)) spanning large distances (20 μ m T cell and macrophage (3)) aggregation and colocalization are necessary mechanisms in the signal transduction pathway. Although the cell membrane is a vastly complex structure

filled with heterogeneous microdomains IFN γ R has been observed to colocalize in caveolar membrane domains (1, 2, 4, 5).

Experimental evidence shows the link between IFN γ and caveolar domains goes beyond spatial organization on the plasma membrane, gene networks of caveolin-1 and IFN γ seem to be entwined. Stimulating macrophages with IFN γ had an inhibitory effect on caveolin-1, the marker protein of caveola (7). In addition when transfecting cell lines HT20 and DLD1 with caveolin-1 cDNA there is downregulation in iNos(6), a metabolic product of IFN γ immune response. A complex regulatory network exists between caveolin-1, IFN γ , and iNos.

This distinct network has yet to be fully elucidated. Starting with a model of the IFN γ immune response (8) we have added gene networks of caveolin-1 as well as gene and metabolic networks of iNOS. In this study our aim is to understand the effects that spatial clustering has on IFN γ R downstream signaling using the coupled spatial non-spatial simulation algorithm (CSNSA) to simulate the IFN γ /IFN γ R mediated activation of the JAK-STAT signal transduction cascade. We then investigate the relationship between iNOS and caveolin-1 using a modeling framework which combines the CSNSA with BioXyce, where BioXyce is used to simulate the STAT mediated intracellular reactions that lead to the production of IFN γ activated genes and the metabolic reactions that lead to the production of immune effector molecules. We discuss the challenges and benefits of the coupled platform in providing a multiscale understanding of host immune response mechanisms.

REFERENCES

- [1] Takaoka, A., et al. (2000) *Science* **288**, 2357-60.
- [2] Sadr, R., et al. (2001) *Cytokine* **14**, 19-26.
- [3] Wada, H., et al. (2008) *Nature* **452**, 768-72.
- [4] Lambert, et al. (2000) *Cytokine* **12**, 715-9.
- [5] Sadr, R., Lortat-Jacob, H. & Morel, G. (2000) *Cytokine* **12**, 711-4.
- [6] Sanchez, F. A., et al. (2008) *Am J Physiol Heart Circ Physiol*
- [7] Felley-Bosco, E. et al. (200) *Proc Natl Acad Sci USA* **97**, 1433-9
- [8] Yamada, S., Shiono, S. Joo, A., and Yoshimura, A. (2003) *FEBS Letters* **534** 190-196
- [9] Schroder, K., Hertzog, P.J., Ravasi, T., Hume, D. (2004) *Jrnl Leukocyte Bio./ 75*, 163-189

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